

CLINICOPATHOLOGICAL STUDIES ON HEPATOPROTECTIVE EFFECT OF FUCOIDAN ON CCL₄ INDUCED LIVER FIBROSIS IN GUINEA PIGS

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ABSTRACT

In this study, we investigate the protective and therapeutic effect of fucoidan extract from Laminaria species against liver damage induced by CCl₄ in Guinea pigs by monitoring the hepatic m-RNA expression of TGFβ-1, some serum biochemical parameters and some oxidative stress biomarker. The CCl₄ was orally administrated at dose 1 ml/kg bw twice a week for 8 weeks. Fucoidan orally administrated at a dose rate 200 mg/kg bw/day. We found that fucoidan treatment improved elevated m-RNA expression of TGFβ-1, T.Bil, D.Bil, Ind.Bil, MDA and SOD serum levels induced by CCl₄ at 8th week post treatment. Meanwhile ALT, AST and AP are not improved by fucoidan treatment. Finally, we concluded that crud fucoidan is less effective as hepatoprotective agent, and had a mild effect in the protective treatment at 8th week post treatment.

INTRODUCTION

Liver diseases are some of the fatal disease in the world today, they pose a serious challenge to international public health. Hepatic fibrosis is a wound healing response to chronic liver injury which characterized by net accumulation of extracellular matrix (ECM) including collagen, glycoproteins and proteoglycan. Hepatic stellate cell (HSC) which previously known as Ito cell that under physiological condition store 80% of retinoids (vitamin A) is the cytological base of the hepatic fibrosis. The quiescent HSC are transformed with progressive injury into myofibroblast like cells that characterized by the appearance of cytoskeleton protein α smooth muscle actin (α SMA) which consider a bio-

marker for HSC activation (Yong et al., 2005). TGFβ-1 is a key molecule or most fibrogenic cytokines facilitate the activation of HSC and convert it from static HSC into phenotype of myofibroblast to express α SMA and posses the character of contraction (Semihha et al., 2006).

Carbon tetrachloride, CCl₄ is a frequently used chemical to experimentally induced hepatic fibrosis. Depending on the dose and duration the effect of CCl₄ on hepatocytes are manifested histologically as hepatic steatosis, fibrosis, hepatocellular death and carcinogenicity (Heekyoung et al., 2005). The hepatotoxic effect of CCl₄ involved in immediate cleavage of CCl₄ by cytochrome P450

(CYP2E1) in hepatocytes which generate trichloromethyl radical leading to lipid peroxidation and membrane damage subsequently. Activated Kupffer cell produce toxic metabolites (inflammatory cytokines and reactive oxygen intermediates) resulting in the injury of hepatic parenchymal cells (Chun et al., 2009).

Fucoidans, is sulfated polysaccharide extracted from cell wall of brown algae and some marine invertebrates. Firstly isolated by Kylin almost one century ago, contain substantial percentages of L-fucose and sulfate ester groups also called fucan, fucosan or sulfated fucan. Recently fucoidan has been extensively studied due to its numerous biological activities including antioagulant, antithrombotic, antitumor, antiviral, anti-complement, antioxidant and anti-inflammatory activities. Also used as immunomodulatory, blood lipid reducing, has activity against hepatopathy, renalpathy and gastric protective effect (Bo et al., 2008). The brown seaweed *Laminaria japonica* is the most important economic seaweed cultured in China. The utilization of *L. japonica* as a drug has been documented in Traditional Chinese Medicine (TCM) for over one thousand years. Fucoidan of *L. japonica* had a hepatoprotective effect (Ning et al., 2005).

This study was aimed to, evaluate hepatoprotective effect of fucoidan on liver fibrosis induced by CCl_4 in guinea pigs through detection of gene expression of TGF β -1 by RT-PCR, oxidative stress reaction through estimation of MDA, SOD, GSH and CAT enzymes, as well as some biochemical parameters.

MATERIAL AND METHODS

Experimental Animals :

Fifty Guinea pigs of 1-2 month old of both sexes were obtained from Helwan farm of laboratory Animals (Ministry of Public health). The animals were kept in galvanized zinc plate cages under strict hygienic conditions. The animals were ensured free from any infection. The Guinea pigs were maintained on pelleted diet and water ad Libitum. The daily requirement of ascorbic acid (50 mg / liter of drinking water) was supplied all over the experiment according to Sarah and Maggie (2003).

Chemicals :

CCl_4 was purchased from ADWIA Co. Egypt., Primer sequences for PCR amplification : Transforming growth factor β 1 (TGF- β 1).

Obtained from metabion international AG., Lend-Christ-Strasse 44 \ 1, Martinsried \ Deutschland.

Gene	Primer sequence		Base pair
TGF- β 1	Sense	5' TAT AGC AAC AAT TCC TGG CG 3'	162
	Antisense	5' TGC TGT CAC AGG AGC AGT G 3'	

Fucoidan :

Fucoidan extract of *Laminaria* species obtained as powder that used as freshly prepared solution dissolved in normal saline. Provided by (Bijing Lilli Agrochemistry CO. LTD, China).

Fibrosis induction and treatment:

Guinea pigs were divided into 5 groups as follow:

Group I (10) served as normal control receive only the vehicle (1ml/kg bw olive oil twice a week for 8 weeks). **Group II (10)** treated with fucoidan (200 mg/kg bw/day all over the experiment) and olive oil. **Group III (15)** receive 1ml/kg bw of CCl₄ diluted 20% in olive oil twice a week for 8 weeks. **Group IV (10)** pretreated with fucoidan for one week before CCl₄ administration then treated with CCl₄ and fucoidan (protective fucoidan treated group). **Group V (5)** after 4 weeks of CCl₄ administration and ensure occurrence of fibrosis (5 animals) further treated with CCl₄ for 4 weeks and at the same time treated with fucoidan (therapeutic fucoidan treated group). At the end of 4th and 8th week of CCl₄ treatment randomly five Guinea pigs were picked up from each group. Blood samples were collected individually from heart puncture for serum chemistry, Guinea pigs were then sacrificed and specimen from liver were kept in liquid nitrogen for reverse transcriptase polymerase chain reaction (RT-PCR) analyses.

RT-PCR analysis:

Total RNA was isolated from Guinea pigs livers using QIAamp extraction method. Preparation of the RNA / primer mix by add the following (RNA template, Forward Primer and

Reverse Primer) to a nuclease-free microtube and mix by pipetting gently up and down. Incubate the mixture at 70-75°C for 5-10 min and place it at room temperature for 5-10 min for denaturation and primer annealing. Prepare the RT-PCR mix and complete it by adding 10 µl RNA/primer mix then pipette on ice and mix by pipetting gently up and down. The thermal profile that was used consisted of denaturation at 95°C, annealing at 55-65°C, elongation at 72°C and Final elongation at 72°C. The PCR product was electrophoresed on 2% agarose gel electrophoresis. RT-PCR analysis was performed in Biotechnology Center for Service and Researches (BCSR), Faculty of Vet. Medicine, Cairo University.

Serum biochemical analysis:

Prepared frozen serum samples were analyzed for alanine aminotransferase (ALT), aspartate aminotransferase (AST), alkaline phosphatase (ALP), total (T. Bili.) and direct bilirubin (D.Bili.), glucose, total protein, albumin (Alb), urica, creatinine (Cre.) and some oxidative stress marker malondialdehyde (MDA) superoxide dismutase (SOD), catalase (CAT) and reduced glutathione (GSH) were determined with semi-automatic spectrophotometer (BM-Germany 5010) using commercial test kits (Randox Co. UK and Biodiagnostic, Egypt.) according to enclosed pamphlets.

Statistical analysis:

Our results were analyzed by (ANOVA) using SPSS software statistical program (SPSS for windows (ver.15.00, USA). Two groups were significantly different if P value was statistically lower than 0.05 .

RESULTS & DISCUSSION

The liver plays a central role in metabolic homeostasis, as it is responsible for the metabolism, synthesis, storage and redistribution of nutrients, carbohydrates, fats and vitamins. Importantly, it is the main detoxifying organ of the body, which removes wastes and xenobiotics by metabolic conversion and biliary excretion (Joan et al., 2010). CCl₄ metabolism is an established model of liver necrosis and fibrosis. The liver damage is created by this metabolism is free radical dependent as CCl₄ is oxidized by cytochrome P450 to highly reactive trichloromethyl (CCl₃) radical that being generated by reductive cleavage of CCl₄ bond and generated oxygen radicals and phospholipid peroxides in abundance (Sreedevi et al., 2006) and (Gopal et al., 2001). The generated trichloromethyl free radical caused liver necrosis, destruction of ECM and lipid peroxidation of membranes. Our study showed that TGF-β1 mRNA expression increased as fibrosis developed in CCl₄ induced liver fibrosis in Guinea pigs (82.68 % and 88.66 at 4th and 8th week respectively). As TGF-β1 activity is enhanced by proteolytic release and activation of latent TGF-β1 by HSC also other cell such as kupfer cells, invading mononuclear cells, myofibroblast cells and endothelial cells can synthesis and release TGF-β1 (Shi-ling et al., 2005).

Fucoidans, a family of sulphated polyfucose polysaccharides, exhibit different biological properties. The biological effects of fucoidan are related to their polysaccharide backbones and sulfate content. Recently the antifibrotic activity of fucoidan was reported in an animal model of hepatic fibrosis

(Kyoumi et al., 2009). The TGF-β1 mRNA expression was reduced by fucoidan treatment especially in (GP.IV) at 8th week post treatment. The TGF-β1 mRNA expression was 77.32% in GP.IV at 4th week and (41.24% and 48.45%) in (GP. IV and GP.V respectively) at 8th week post treatment with CCl₄ and fucoidan. This is agreeing with Shinji et al., (2008) who mentioned that fucoidan treatment attenuate HSC activation by inhibiting TGF-β1.

CCl₄ administration, causes severe liver damage demonstrated by remarkable elevation of serum AST and ALT levels till the end of the experiment as shown in (table.1,2). This elevation may be attributed to the cellular leakage and damaged of structural integrity of the liver cells (Robert et al., 2009). Also CCl₄ treatment induced elevation of serum ALP with high level of total bilirubin, direct and indirect bilirubin (table.1,2). Which are considered indicator of cholestasis and pathological alterations of the biliary flow (Lalitstingh et al., 2010). The high concentration of indirect bilirubin and direct bilirubin in the serum is an indication for increased erythrocyte degeneration rate and liver injury caused by CCl₄ (Sathesh et al., 2009).

In our work fucoidan not have any toxic effect either on liver or kidney function as ALT, AST, ALP, BUN and creatinine level are in normal level in GP.II (table.1,2). The elevated liver enzyme (ALT, AST & ALP) induced by CCl₄ not improved by fucoidan treatment either at 4th or 8th week either in protective (GP.IV) or therapeutic treatment (GP.V) (table.1,2). This is in accordance with Kyoumi et al., (2009) who observed that crude fucoidan

extract not improved elevated ALT and AST serum levels in liver injury induced by N-nitrosodiethylamine and concluded that crude fucoidan showed unremarkable anti-fibrogenesis activity. On other hand **Shinji et al., (2008)** reported that administrated fucoidan by I/V injection in chronic liver injury induced by CCl₄ in mice reduce elevation in serum AST & ALT. The differences may be due to molecular weight of fucoidan that was reported in the anti-tumor activity, pro-angiogenesis and antioxidant activities of fucoidan and route of administration as anti-fibrogenic effect of fucoidan by oral administration not studied (**Kyoumi et al., 2009**). The T.Bil. (table.1,2) improved by fucoidan treatment either protective or therapeutic one but Ind. Bil. return to normal level in PF group could be due to decrease erythrocyte damage induced by CCl₄ as fucoidan has strong antioxidant activity (**Bo et al., 2008**). In the present study, serum glucose (table.1,2) was reduced in CCl₄ treated animal along of the experiment as hepatic glycogen content was decreased reflecting decreased gluconeogenesis by the liver (**Omar et al., 2007**). In addition, **Rui et al., (2002)** recorded that gluconeogenesis and Krebs cycle fluxes are altered in rat livers following CCl₄ intoxication. Meanwhile **Bhatia and Ahujara, (1984)** suggested that hypoglycemia may be due to depletion of glucose-6-phosphatase in the liver of CCl₄ treated animal. Fucoidan treatment not relieve hypoglycemia induced by CCl₄ (table.1,2). TP and albumin blood level insignificantly changed by CCl₄ treatment except at 8th week post treatment albumin level was increased (table.1,2). This is may be due to dehydration as a result of diarrhea caused by CCl₄ treatment. Globulin level in our work

was increased by CCl₄ treatment at 4th week but decreased at 8th week post treatment. The increased globulin level occurred due to inflammation and liver disease. Meanwhile CCl₄ significantly decrease globulin at 8th week post treatment which attributed to reduce immunoglobulin IgM, IgA and specially IgG by CCl₄ in a dose and time dependant manner (**Usama, 2009**). In fucoidan treated groups (GP. II, IV & V) total protein, albumin, globulin and A/G ratio insignificantly change in comparing with control (table.1,2). Except globulin level increase in (GP.II) at 4th week post treatment. Fucoidan has both humoral and cell-mediated immune responses as it enhance B cell blastogenesis. Therefore, the fucoidan was expected to promote maturation of B cells that might result in the stimulation of antibody secreting activity (**Kyoko et al., 2008**). Presently administration of CCl₄ to normal Guinea pigs induced renal toxicity revealed by elevation of urea at 4th week post treatment and creatinine level which occurred at 8th week post treatment only (table.1,2). As CCl₄ administration mediated peroxidation of lipid structures and protein content of renal tissues, resulting in sub cellular damages (**Muhammad et al., 2009**). Malondialdehyde is a reactive aldehyde, used as an indicator of the amount of lipid peroxidation (**Shana et al., 2009**). In the present study, the significant increase in serum MDA concentration observed in the CCl₄ treated group (GP.III) as Lipid peroxidation (LPO), is accepted to be one of the principal causes of CCl₄ induced liver injury, the reaction resulting from the attack by reactive free radicals on the polyunsaturated fatty acids (PUFAs) to generate different products, including volatile alkanes, aldehydic products that are relatively stable resulting

ultimately in a loss in membrane integrity (Robert et al., 2009).

In fucoidan treated groups (GP.IV & V) MDA (table.3,4) elevated at 4th week post treatment but insignificantly change at 8th week post treatment in comparing with both CCl₄ and control one. This agrees with Bo et al., (2008) who reported that fucoidan from *L.japonica* had no effect on lipid peroxidation induced by FeSO₄ in vitro. Also Kyoumi et al., (2009) mentioned that crude fucoidan extract not reduced high MDA level in liver injury induced by N-nitrosodiethylamine. In contrast Kum et al, (2008) found that I/P administration of fucoidan extract resulted in reduced high MDA level induced by CCl₄ treatment in rats. Antioxidant enzyme such as SOD, CAT and GSH constitute a supportive team of defense against ROS. Our result

found an elevation in SOD, CAT and GSH in Guinea pigs treated with CCl₄ at 4th week (table.1,2). The increase in these enzyme activities was probably a response towards the increase in ROS generation (Gowri et al., 2008). The fucoidan treatment correct SOD level in protective group as fucoidan has strong scavenging free radical activity especially against superoxide radical. This is agreeing with Jing et al., (2008) who mentioned that fucoidan exhibit radical scavenging activity in vitro and antioxidative activity against oxidative stress in cellular model. Finally we concluded that crude fucoidan is less effective as hepatoprotective agent, and it had mild prophylactic hepatoprotective effect at 8th week post treatment. Further research recommended for more definitive knowledge about efficacy of fucoidan as hepatoprotective agent.

Table (1, 2): Some Serum Biochemical Profiles and Oxidative Stress Biomarkers (Mean ± S.E) at 4th (table, 1) and 8th (table,2) Week Post Treatment with CCL₄ and Fucoidan in Guinea pigs.

(table,1)

Groups	ALT U/L	AST U/L	ALP U/L	T. Bil. mg/dl	Dir. Bil. mg/dl	Indir. Bil. mg/dl	Glucose mg/dl	TP gm/dl	Alb. gm/dl	Glob. gm/dl	A/G	Urea mg/dl	Cre. mg/dl
I (Cont)	20.64 ^a ±1.30	29.11 ^a ±1.62	10.92 ^a ±0.67	0.47 ^a ±0.03	0.22 ^a ±0.02	0.25 ±0.04	134.80 ^a ±2.20	4.58 ±0.23	3.21 ±0.30	1.37 ^a ±0.11	2.49 ^b ±0.43	55.89 ^a ±1.99	0.58 ±0.01
II (F)	19.78 ^a ±1.81	28.00 ^a ±1.34	9.52 ^a ±0.51	0.50 ^{ab} ±0.05	0.25 ^a ±0.06	0.25 ±0.05	134.60 ^a ±4.52	5.83 ±0.48	3.17 ±0.29	2.67 ^b ±0.35	1.28 ^a ±0.16	60.27 ^{ab} ±1.12	0.61 ±0.05
III (CCL ₄)	34.19 ^b ±2.03	39.70 ^b ±1.46	16.40 ^b ±0.60	0.72 ^{bc} ±0.1	0.53 ^b ±0.12	0.19 ±0.03	97.20 ^a ±8.53	5.59 ±0.74	2.99 ±0.25	2.59 ^a ±0.74	1.44 ^a ±0.27	71.55 ^c ±2.09	0.67 ±0.06
IV (PF)	33.29 ^b ±2.09	37.30 ^b ±2.54	16 ^b ±0.93	0.75 ^c ±0.09	0.52 ^b ±0.09	0.23 ±0.03	96.60 ^a ±10.54	5.24 ±0.39	3.29 ±0.27	1.95 ^{ab} ±0.24	1.78 ^{ab} ±0.26	66.67 ^{bc} ±3.23	0.66 ±0.02

(table,2)

Groups	ALT U/L	AST U/L	ALP U/L	T. Bil. mg/dl	Dir. Bil. mg/dl	Indir. Bil. mg/dl	Glucose mg/dl	TP gm/dl	Alb. gm/dl	Glob. gm/dl	A/G	Urea mg/dl	Cre. mg/dl
I (Cont)	19.93 ^a ±0.07	30.98 ^a ±1.20	9.99 ^a ±0.41	0.45 ^a ±0.02	0.24 ^a ±0.03	0.21 ^a ±0.02	133.65 ^a ±2.52	4.76 ±0.13	2.98 ^a ±0.22	1.78 ^a ±0.21	1.86 ^a ±0.35	57.36 ±3.76	0.54 ^a ±0.01
II (F)	20.63 ^a ±0.86	29.89 ^a ±1.61	9.75 ^a ±0.45	0.47 ^a ±0.02	0.17 ^a ±0.03	0.30 ^{ab} ±0.04	133 ^b ±2.85	5.07 ±0.11	3.35 ^a ±0.13	1.71 ^b ±0.17	2.08 ^a ±0.26	56.92 ±4.24	0.59 ^{ab} ±0.02
III (CCL ₄)	41.76 ^b ±4.19	41.07 ^b ±0.65	16.93 ^b ±0.53	0.80 ^b ±0.06	0.42 ^b ±0.08	0.38 ^b ±0.08	95.60 ^a ±13.57	4.9 ±0.11	3.92 ^b ±0.22	0.98 ^a ±0.15	4.47 ^b ±0.85	59.30 ±7.17	0.71 ^c ±0.02
IV (PF)	36.96 ^b ±2.51	37.60 ^b ±4.09	18.05 ^b ±2.04	0.53 ^a ±0.02	0.31 ^{ab} ±0.02	0.22 ^a ±0.02	106.40 ^a ±10.06	4.98 ±0.13	3.30 ^a ±0.19	1.68 ^a ±0.28	2.31 ^a ±0.54	59.95 ±9.32	0.73 ^c ±0.04
V (TF)	37.61 ^b ±4.49	37.89 ^b ±2.61	15.84 ^b ±1.94	0.49 ^a ±0.08	0.24 ^a ±0.09	0.25 ^{ab} ±0.09	93.25 ^a ±9.92	4.91 ±0.10	3.21 ^a ±0.18	1.69 ^a ±0.08	1.95 ^a ±0.19	56.17 ±2.68	0.65 ^{ab} ±0.06

Cont. (control), F (fucoidan alone), CCL₄ (carbon tetrachloride treatment), PF (protective fucoidan treatment) TF (therapeutic fucoidan)
The same column not followed by the same letter differ significantly (P<0.05).

Table (3,4): Some Serum Oxidative Stress Biomarkers (Mean \pm S.E) at 4th (table,3) and 8th (table,4) Week Post Treatment with CCl₄, and Fucoidan in Guinea pigs.

(table,3)

Groups	MDA nmol/ml	SOD U/ml	CAT U/L	GSH mg/dl
I (Cont)	7.05 ^a ± 0.09	755.60 ^a ± 41.01	311.25 ^a ± 65.36	0.53 ^a ± 0.08
II (F)	8.52 ^{ab} ± 0.64	882.81 ^{ab} ± 104.51	431.25 ^{ab} ± 22.66	0.58 ^a ± 0.06
III (CCl ₄)	12.45 ^c ± 1.62	947.88 ^b ± 59.67	572.65 ^c ± 57.58	0.77 ^b ± 0.03
IV (PF)	10.93 ^{bc} ± 0.85	725.26 ^a ± 26.06	459.98 ^{bc} ± 36.99	0.87 ^b ± 0.06

(table,4)

Groups	MDA nmol/ml	SOD U/ml	CAT U/L	GSH mg/dl
I (Cont)	7 ^a ± 0.09	744.04 ^{ac} ± 39.58	403.65 ^{ab} ± 59.59	0.55 ^a ± 0.06
II (F)	7.03 ^a ± 1.14	638.81 ^a ± 8.23	393.01 ^a ± 37.53	0.59 ^a ± 0.19
III (CCl ₄)	11.30 ^b ± 1.79	715.89 ^{ab} ± 59.51	502.48 ^{bc} ± 40.12	1.49 ^b ± 0.18
IV (PF)	9.70 ^{ab} ± 2.19	854.60 ^c ± 73.78	448.72 ^{ac} ± 49.19	1.21 ^b ± 0.20
V (TF)	10.50 ^{ab} ± 1.29	798.36 ^{cb} ± 15.65	547.58 ^c ± 40.31	1.25 ^a ± 0.06

Cont.(control), F (fucoidan alone), CCl₄ (carbon tetrachloride treatment), PF (protective fucoidan treatment) TF (therapeutic fucoidan).

The same column not followed by the same letter differ significantly (P<0.05).

REFERENCES

- Bhatia, B. and Ahujara, P. L. (1984)** : Cold tolerance in CCl₄ treated rats and its modification by administration of garlic oil and glucose. *Int J Biometeor* ;28 (2):93-99.
- Bo, L.; Fei, L.; Xinjun, W. and Rutxdang, Z. (2008)** : Fucoidan, structure and bioactivity. *Molecules*; 13:1671-1695.
- Chun-Ping, C.; Ping, W.; Yang, L.; Da-Jin, Z.; Li-Sheng, W. and Chu-Tse, W. (2009)** : The protective role of hepatopoietin Cn on liver injury induced by carbon tetrachloride in rats. *Hepatology Research*; 39 (2):200-206.
- Gopal, M. K.; Kathleen, H. A.; Jackson, R. L.; Jason, M. D.; Robert, M. C.; Larry, T. W.; Stephen, P. M.; Guy, A. Z. and Thomas, M. M. (2001)** : Identification of platelet activating factor as the inflammatory lipid mediator in CCl₄-metabolizing rat liver. *Journal of Lipid Research*; 42(4):587-596.
- Gowri, S.; Manavalan, R.; Venkappayya, D. and David, R. (2008)** : Hepatoprotective and antioxidant effects of Commiphora berry (Arn) Engl bark extract against CCl₄ induced oxidative damage in rats. *Food and Chemical Toxicology*; 46(9): 3182-3185.
- Heekyoung, C.; Doo-Pyo, H.; Ji-Youn, J.; Hyun-Jun, K.; Ki-Seok, J.; Yhun-Yhong, S.; Joon-Ik, A.; Yong-Sung, L. and Gu, K. (2005)** : Comprehensive analysis of differential gene expression profiles on carbon tetrachloride-induced rat liver injury and regeneration. *Toxicology and Applied Pharmacology*; 206(1):27- 42.
- Jing, W.; Quanbin, Z.; Zhongshan, Z. and Zhen, L. (2008)** : Antioxidant activity of sulfated polysaccharide extracted from *Laminaria japonica*. *International Journal of Biological Macromolecules*; 42(2): 127-132.
- Joan, O.; Barbara, A. F.; Qing, X. and Samuel, W. F. (2010)** : The identification of stem cells in human liver diseases and hepatocellular carcinoma. *Experimental and Molecular Pathology* ; 88(3):331-340.
- Kum, S. K.; In, D. K.; Ryun, H. K.; Jin, Y. L.; Jac, S. K. and Bac, J. H. (2008)** : The Effects of Fucoidan Extracts on CCl₄-Induced Liver Injury. *Arch Pharm Res*; 31(5): 622-627
- Kyoko, H.; Takahisa, N.; Minoru, H.; Kenji, K. and Toshimitsu, H. (2008)** : Defensive effects of a fucoidan from brown alga *Undaria pinnatifida* against herpes simplex virus infection. *International Immunopharmacology*; 8(1):109-116.
- Kyoumi, N.; Hisashi, T.; Masahiko, I. and Takeaki, N. (2009)** : Attenuation of Nitrosodiethylamine-induced liver fibrosis by high molecular weight fucoidan derived from *C. okamuranu*. *Journal of Gastroenterology and Hepatology*. Article in press.
- Lalitsingh, R.; Jigar, B. and Jagruti, P. (2010)** : Hepatoprotective activity of ethanolic extract of bark of *Zanthoxylum armatum* DC in CCl₄ induced hepatic damage in rats. *Journal of Ethnopharmacology*;127(3):777-780.
- Muhammad, R. K.; Wajtha, R.; Gul, N. K.; Rahmat, A. K. and Saima, S. (2009)** : Carbon tetrachloride-induced nephrotoxicity in rats: Protective role of *Digera muricata*. *Journal of Ethnopharmacology*; 122(1) : 91-99.
- Ning, L.; Quanbin, Z. and Jinming, S. (2005)**: Toxicological evaluation of fucoidan extracted from *Laminaria japonica* in Wistar rats. *Food and Chemical Toxicology*; 43 (3):421-426.
- Omar, A. S. M. E.; Amany, S. A.; Enayat, O. A. and Nabila, H. S. (2007)** : Effect of

ribavirin alone or combined with silymarin on carbon tetrachloride induced hepatic damage in rats. *Drug Target Insights*; 2: 19-27.

Robert, D.; Hrvoje, J.; Cedomila, M. and Biserka, R. (2009) : Dose- and time-dependent effects of luteolin on carbon tetrachloride-induced hepatotoxicity in mice. *Experimental and Toxicologic Pathology*; 61 (6):581-589.

Rui, A. C.; John, G. J.; Chris, M. G.; Dean, A. S. and Craig, R. M. (2002) : Hepatic gluconeogenesis and krebs cycle fluxes in a CCl4 model of acute liver failure. *NMR Biomed*;15(1): 45-51.

Sarah, W. and Maggie, L. (2003): Handbook of Laboratory Animal Management and Welfare.3rd ed. USA at Iowa State Press.

Sathesh, K. S.; Ravi, K. B. and Krishna, M. G. (2009) : Hepatoprotective effect of *Trichosanthes cueumerina* Var *eucumerina* L. on carbon tetrachloride induced liver damage in rats. *Journal of Ethnopharmacology*; 123 (2): 347-350.

Semiha, N.; Ukin, C. and Zehra, M. F. (2006): The effect of vitamin A on CCl4-induced hepatic injuries in rats: a histochemical, immunohistochemical and ultrastructural study. *Acta Histochemica*; 107(6):421-434.

Shana, R. D.; Serene, M. L.; Rachel, N. K.; Amin, A. N.; Kusum, K. K.; Carol, A. C. and Benita, L. M. (2009) : Carbon tetrachloride-induced liver damage in alloxan-

glycoprotein receptor-deficient mice. *Biochemical pharmacology*; 77(7):1283-90.

Shi-Ling, S.; Zuo-Jiong, G.; Quan-Rang, Z. and Tuan-Xin, H. (2005) : Effect of Chinese traditional compound, JinSanE on expression of TGF- β 1 and TGF- β 1 type II receptor mRNA, Smad 3 and Smad 7 on experimental hepatic fibrosis in vivo. *World J Gastroenterol*; 11(15):2269-2276.

Shinji, H.; Ayano, I.; Katsuhiko, I.; Masuo, K.; Masaya, K. and Kiyohito, Y. (2008) : Fucoidan partly prevents CCl4-induced liver fibrosis. *Eur J Pharmacol*; 580 (3): 380-384.

Sreedevi, A.; Lie, Y.; Yan, S.; Alice, W. L.; Wood-Yee, C.; Wing-Tai, C.; Wing-Tai, C. and Susanna, S. L. (2006) : A temporal study on the histological, biochemical and molecular responses of CCl4 induced hepatotoxicity in Cyp2e1-null mice. *Toxicology*; 228 (2-3):310-322.

Usama, B. E.; Ali, H. E. and Hussein, A. A. (2009) : The ameliorative effect of phoenix dactylifera extract on CCl4 hepatotoxicity in new zealand rabbits. *Journal of Applied Sciences Research*; 5(9): 1082-1087.

Yong, H. X.; Dian, W. L. and Qing, L. (2005) : Effect of drug serum of anti-fibrosis 1 herbal compound on calcium in hepatic stellate cell and its molecular mechanism. *World Journal of Gastroenterology*;11(10):1515-1520.

الملخص العربى

دراسات باثولوجية إكلينيكية على الفيوكيدان كعامل وقائى للتليف الكبد المحدث برابع كلوريد الكربون فى خنازير غينيا

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ط. ب. فاطمه مصطفى عبد الحميد*

قسم الباثولوجيا الإكلينيكية - كلية الطب البيطرى - جامعة المنصورة*

قسم الباثولوجيا - كلية الطب البيطرى - جامعة المنصورة**

أجريت هذه الدراسة على عدد من خنازير غينيا بهدف تقييم مدى فاعلية الفيوكيدان كعامل وقائى لمرض التليف الكبدى المحدث برابع كلوريد الكربون من خلال الفحص الجينى لأنسجة الكبد لتحديد كمية جين ترانسفورمينج - 1 بالإضافة إلى تحديد الاختلافات فى المؤشرات الكيميائية ودلالات الإجهاد التأكسدى، بعد 4 أسابيع من العلاج بالفيوكيدان لم يظهر أى تأثير للدواء على مكونات السيرم الكيميائية ولا على أكسدة الدهون ومضادات الأكسدة ولكن انخفض Gene expression of TGF β -1 بعد 8 أسابيع من العلاج ظهر له تأثير جزئى من خلال فحص بعض المركبات الكيميائية وخاصة فى المجموعة الوقائية وأدى إلى تقليل كمية جين ترانسفورمينج - 1.